Induced pluripotent stem cells (iPSCs) based approaches for hematopoietic cancer therapy

Bardia Khandany BSc¹, Mohammad Mehdi Heidari PhD¹,*, Mehri Khatami PhD¹

1. Department of Biology, Faculty of Science, Yazd University, Yazd, Iran.

*Corresponding author: Dr Mohammad Mehdi Heidari, PhD, Department of Biology, Faculty of Science, Yazd University, Pajoohesh Street, Yazd, Iran. Email: heidarimm@yazd.ac.ir.

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Abstract

Induced pluripotent stem cells (iPSCs) are reprogrammed from somatic cells through numerous transcription factors. Human induced pluripotent stem cell approaches are developing as a hopeful strategy to improve our knowledge of genetic association studies and the underlying molecular mechanisms. Rapid progression in stem cell therapy and cell reprogramming provides compelling reasons for its feasibility for treating a wide range of diseases through the replacement of autologous cells. Continuous failure in embryonic stem cells (ESC) production and the dependency of iPSC on ectopic genes may be due to the inability to maintain the stability of the endogenous gene systems which are essential for creation of pluripotency state. With recent developments in the genome processing and human tissue culturing approaches as well as xenotransplantation, bioengineering, and genome editing, induced pluripotent stem cells offer the new opportunities for the study of human cancers. Most hematopoietic malignancies are originated from cells that are functionally heterogeneous and few of them are responsible for maintaining tumor state. The naming of these cancer stem cells are due to the quality characteristics of normal tissue stem cells, such as self-renewal, long term survival, and the ability to produce cells with more differentiated properties. The aim of present study was to focus on the recent progresses in the application of stem cell-based hematopoietic cancer, and to assess the benefits of treatment, opportunities, and shortcomings that can potentially help improve future efforts in experimental and clinical studies.

Keywords: Gene therapy, Hematopoietic cancer, Induced pluripotent stem cells

Introduction

The human body comprises of over two hundred different types of cells that form tissues and organs and provide all the essential functions for survival reproduction (1). All of cells in the body, both somatic (of all three germ layers) and germ cells, originate from the embryonic pluripotent cells (2). In 1961, for the first time, Till and McCulloh (3) introduced self-renewal properties in any living cells as one of their important breakthrough. They influenced the mice with lethal radiation doses and then injected bone marrow cells in mice. They found that the main reason of survived of the mice was these cells formed clumps due to cell cloned from them (4, 5). In 2006, Japanese researchers at Kyoto University recognized conditions that allowed specialized adult cells to be genetically "reprogrammed" to accept a stem cell-like state (6). These specialized cells, which are now called induced pluripotent stem cells (iPSCs), , were reprogrammed to an embryonic stem cell-like state via introducing important genes to maintaining the critical properties of embryonic stem cells (ESCs) (7). After this discovery, researchers have improved the generation techniques of iPSCs and created powerful new pathways to dedifferentiate cells, which their growth and developmental conditions was already determined (8-10). The main focus of worldwide investigators and clinicians is on the potential use of iPSCs as a helpful disease for modeling, development, and regenerative medicine

(11-13). Furthermore, the ethical issues related to the ESCs generation are not applied to iPSCs, proposing a non-controversial strategy to create pluripotent cells using existing non-pluripotent cell lines which can be transplanted into the patients without the concern of immune rejection (autologous graft) (14, 15). However, there are many challenges in pre-temporary iPSC tests, one of which is teratoma development or tumorigenesis risk if they are inserted in sensitive sites such as promoters and host gene enhancers (16).

Experimental strategies for iPSCs generation

One strategy to achieve iPSCs generation is nuclear reprogramming, a stable alteration in the nucleus of a mature cell, which can then be preserved and replicated as the cell divides through mitosis (17). strategy is implemented using This techniques such as fuse somatic cells with ESCs, somatic cell nuclear transfer (SCNT), and altered nuclear transfer (ANT) (18,19). The nuclear reprogramming approach involves the use of mature "somatic" cells from an adult and the introduction of genes that encoding critical transcription factor proteins that themselves regulate the function of other important genes in the early developmental stages of the fetus (20, 21).

In the previous studies, it was found that only four transcription factors (Oct4: Octamer binding transcription factor-4, Sox2: Sex determining region Y-box2, Klf4: Krupple like factor 4, and c-Myc) were required for reprogramming of mouse fibroblasts to an embryonic stem cell-like state by forcing them to express important genes for maintenance of the defining properties of ESCs (22-24). In 2007, two different research teams reached a new milestone through deriving iPSCs from human cells and using the original four genes containing Oct4, Sox2, Nanog, and Lin28 (25, 26). Since then, researchers have achieved the production of iPSCs

from somatic tissues of the rat and monkey (27-29).

Several methods have been considered for improving the effectiveness reprogramming and reducing the potentially harmful side effects of the reprogramming process (30). Since the retroviruses used to deliver the four transcription factors in the earliest studies can be potentially mutagens, researchers are not certain about whether all four factors are absolutely essential (23, 31, 32). Specially, the c-Myc gene is known to increase the rate of tumor growth in some cases, which negatively affects iPSC efficiency in transplantation treatments (27, 33). For this purpose, researchers proposed a three-factor approach using Oct4 with the orphan nuclear receptor Esrrb and Sox2, and they were able to transform mouse embryonic fibroblasts into iPSCs. Similarly, other reports have indicated that c-Myc has not a functional role for direct reprogramming of mouse fibroblasts (34, 35). Studies have further reduced the number of essential genes for reprogramming, and investigators continue to recognize of chemicals that can replace or increase the efficiency of transcription factors in this procedure (36, 37). These successes continue to inform and facilitate reprogramming process, advancing the field toward the production of patient specific stem cells for clinical application. However, the way through which transcription factors transfer to the somatic cells is critical to their potential use in the clinic (38).

IPSCs in cancer therapy: opportunities and challenges

Normally derived iPSCs from patient tissues can mainly be employed for tumor regeneration or treatment of the affected In regenerative medicine, tissues. numerous tissues can be produced by using iPSCs (2, 39). IPSCs therapy may be alternative for damaging good approaches such chemotherapy, as radiotherapy, or surgical treatment.

However, regenerative therapy mediated by human iPSCs needs strong in vivo evidence from iPSC-derived tissues (19, 40).

Now, only a few types of human iPSCderived cells (e.g. hepatocytes) have been effectively assessed in vivo (41, 42). In addition to the direct role of iPSCs in cancer treatment, they can be utilized to screen novel anticancer drugs. Derived iPSCs from differentiating cancer tissue generate cell types that may be more biologically linked to human tumors and be more suitable for drug screening methods. IPSCs are also better candidates than other stem cells for evaluation of the toxicities of antitumor drugs (25, 37, 43). chemotherapy Similar to most components, stem cell therapy using a single agent commonly cannot eliminate tumors. Therefore, a desired combination should be rationally generated and selected (44, 45). Many of these combination therapies have been tested to improve treatment durability. For example, chemotherapy combined with interferon (IFN)-beta immunotherapy, by using a pro-drug/suicide gene system, has displayed synergistic therapeutic effects in human colorectal cancer (44, 46). In 2011, Zielske et al., found that irradiating tumor cells could induce production of factors that stimulate stem cell invasion and increase the number of stem cell in tumors

There are three possibilities for tumors in which cancer stem cells play a critical role. First, the mutations in the normal stem cells or progenitor cells and transformation of them into cancerous stem cells which can lead to the growth of the primary tumor. Second, many of the primary tumor cells may be killed during chemotherapy, but if the cancerous stem cells are not eliminated, they become cancer-resistant stem cells and may result in tumor recurrence. Third, the cancer stem cells may emigrate to distal positions from the primary tumor and cause metastasis (48). For these reasons, most previous

researchers paid special attention to the pluripotent stem cells that might be highly tumorigenic. However, recently it is shown that iPSCs is safer for clinical use than ESCs and other stem cells (49). However, various strategies are used to minimize the risk and possibility of neoplastic transformation.

undifferentiated In the first step, pluripotent stem cells, which potentially tumorigenic, can be separated from clinical approaches using antibodies that focus on specific surface-displayed biomarkers. Stem cell differentiation ofdownregulates expression these biomarkers (50).

Second, directed differentiation of iPSCs involves monitoring of the expression of specific differentiation Successfully differentiated cells can be recognized and screened by using recombinant reporter proteins. Green fluorescent protein (GFP) and similar proteins work well as reporters of against undifferentiated differentiated cells (51). Third, antibody-guided toxins or antibodies can kill the undifferentiated pluripotent stem cells through immune pathways. For instance, monoclonal antibodies against claudin-6, a surface biomarker for undifferentiated pluripotent ESCs and iPSCs, can guide immune-toxins to these stem cells for targeted and selective killing (52).

Fourth, undifferentiated pluripotent stem cells can be removed from cytotoxic components, which can be utilized to kill targeted and selectively pluripotent stem cells that can progress into tumors (53).

Fifth, potentially tumorigenic pluripotent stem cells can be transformed with suicide genes for sensitization towards pro-drugs. For this purpose, the enzyme/pro-drug cancer therapy strategies can be improved to kill undifferentiated pluripotent stem cells (54) Finally, tumorigenic pluripotent stem cells can be eradicated through self-induced transgenic expression of recombinant human DNases (55). These

(47).

strategies can protect a large population of stem cells against tumor transformation.

Application of iPSCs in regenerative medicine for cancer therapy

It is proven that reprogramming of human primary cancer cells is very difficult. However, there have been relatively few reports demonstrating iPSCs production for successful reprogramming of tumoric human cells. The generation of novel iPSC and reprogramming cancer cells are thought to be faced with difficulties such reprogramming-triggered cellular senescence, tumor-specific genetic mutations. epigenetic modifications, genomic instability, and accumulation of DNA damage. In spite of these difficulties, several researchers have reported the generation of new iPSC lines from present cancer cell lines. Reports of iPSC lines resulting from human cancer cell lines are summarized in Table I, which presents a range of cancers such as leukemia (AML) (56), breast cancer (57), retinoblastoma (58), colon cancer (44), Lliver cancer (59), hematologic and solid malignancies (60), melanoma (61), myelomonocytic leukemia (62), and glioblastoma (63). Based on these studies, reprogramming of cells with human cancer cell lines is not an impossible task. However, certain types of cancers may be commonly used against reprogramming factors and combination of factors should be carefully selected depending on the type of cancer cells that are attempting to reprogram. Nevertheless, reports of iPSCs generated from human primary malignant cells are few and limited to certain cancers, especially leukemia. Hu et al., were successful in reprogramming primary human lymphoblasts using transgene-free iPSC technology to ectopically express OSKM, LIN28, NANOG, and the SV40 large T gene from a BCR⁻ABL⁺ CML

Kim et al., patient (64). reported generation of single iPSC-like line in the parental pancreatic ductal adenocarcinoma (PDAC) cancer cultures that contained KRAS G12D mutation (65). The rarity of successful studies demonstrates difficulty of reprogramming primary cancer cells to iPSCs. However, technical limitations, such as complications in maintenance primary cancer tissues in culture, cannot be excluded; furthermore, the basic biological barriers may directly undermine the reprogramming process in cancer cells.

In addition to particular characteristic of pluripotent stem cells, such as their selfrenewal and differentiation capabilities, they can repair human tissues after chemotherapy (66). Clinically, after treatment of malignancies with high-dose radiotherapy or chemotherapy, transplanting human stem cells has been generally used to facilitate lifelong hematological recovery (67). The aim of this treatment is to reconstitute the bone marrow under conditions of marrow failure (for example, in aplastic anemia or other blood cell genetic diseases) and to apply human stem cells that differentiate into a desired type of hematopoietic cells in recipients (68). Normally originated iPSCs from patient tissues theoretically be utilized to regenerate tumor- or treatment- damaged tissues (69). In regenerative medicine approaches, different tissues can be produced using iPSCs. IPSC therapy in regenerative medicine may be valuable in replacing or repairing cancer damaged cell radiotherapy, chemotherapy, or surgical treatment (70). However, regenerative therapy mediated by human iPSCs needs in vivo extensive studies in iPSC-derived tissues. So far, only a small number of human iPSC-derived cells, such as hepatocytes, have been successfully evaluated in vivo (71, 72).

		Table I: 1	<i>Applic</i>	ations	of iPSC	s in ca	ncer therap	рy
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	Table I: Applications of		
Origin cells used for generation of iPSC	Reprogramming factors	Cancer type	Type of therapy
Hematopoietic cells (Primary AML cells)	OSKM	Leukemia	investigating the mechanistic basis and clonal properties of human AML, leukemic DNA methylation/gene expression patterns
BRCA1-deleted blood cells	OSKM	breast cancer	Cancer development modeling
Skin cells from RB patient	OSNL	Retinoblastoma	Cancer development modeling
Familial adenomatous polyposis (Colonic organoids)	OSKM	Colon cancer	Modeling of drug Testing in Colorectal Cancer
Hepatic cells	OSKM	Liver cancer	The engraft the liver in a mouse transplantation model
Natural killer cells (BJ1- iPS12, UCBiPS7, and DRiPS16)	OKSM	hematologic and solid malignancies	Anti-tumor therapy (tumor targeting)
human tumor antigen- specific T cells (CD8+ mature T-cells)	OSKM	melanoma	Anti-tumor therapy(tumor targeting)
Dendritic cells (cell lines: 201B7, 253G4, CIRA188Ai-W2, and CB-A11)	OSKM, BMP4	immunological disorders	Cancer immunotherapy such as DC-based vaccines
Juvenile Myelomonocytic Leukemia (JMML)	OKSM	Myelomonocyti c Leukemia	Cancer development modeling
glioblastoma (neural lineage)	OSKM, Oct4	glioblastoma	Cancer development modeling

IPSCs and hematopoietic cancer therapy

Developments in nuclear reprogramming of somatic cells lead to new ways of creating pluripotent stem cell lines and represent a significant step in the production of stem cell-based specific therapies (73).Pluripotency reprogramming approach is based on transfer of a somatic cell nucleus into an

enucleated oocyte, and applying the expression of specific reprogramming genes in somatic cells (74, 75). These two methods are used to reprogram multiple various cell types, including pancreatic β cells, liver cells, fibroblasts, T and B lymphocyte cells, and neural progenitor cells (76-78). Although many types of cells can be under nuclear reprogramming, the fundamental differences between pluripotent stem cell lines derived from distinct somatic cell types are largely unclear.

An immune-mediated antitumor effect resulting from allogeneic hematopoietic stem cell (HSC) transplantation might be appropriate for the treatment of some hematological malignancies, such as CML, ALL, AML, Hodgkin and Non-Hodgkin lymphoma, multiple myeloma (MM), sickle cell anemia, thalassemia, Fanconi anemia (79-81). These HSCs are the most common multipotent stem cells that can differentiate to all blood cell lines through hematopoiesis process (82, 83). After Yamanaka studies in the field of reprogramming, which led invention of iPS technology, many researchers have tried to generate induced hematopoietic stem cells (iHSC) derived from patients blood malignancy (84, 85). with Additionally, these researchers have shown that the specific expression of 13 transcription factors such as OCT4 of OKSM factors can generate HSC. Recently, it has been proven that new genetic engineering tools can be used for producing of iHSCs, including CRISPRmeganucleases, zinc-finger Cas9, nucleases (ZFN), double-strand break nucleases, and transcription (DSB) activator-like effector nucleases (TALEN) (86, 87). Studies have revealed that two transcription factors (HOXA and ERG) act as the core proliferation and differentiation factors in HSC and are inducers of the selfrenewal ability, and thus dysregulation of these factors progresses several types of leukemic cells (85). The main differential marker of hematopoietic stem cells is CD34 that help distinguish iHSC in association with other differentiation markers such as CD34, CD38, CD45, CD90, CD105, CD133, and C-kit (stem cell receptor) (83).

1- Reprogrammed T-lymphocytes

Presenting genes encoding chimeric antigen receptors (CARs) or T-cell receptors (TCRs) directed against tumorrelated antigens makes HSCs attractive for researches of cancer immunotherapy (88-90). Patient-specific iPSCs can be also potentially helpful in immunotherapy methods (91, 92). The pre-rearranged TCR gene is reserved in T lymphocyte-derived human iPSCs, which can be further differentiated into functionally active T cells (93, 94).

In recent years, various technologies, generally based on reversing immunosuppression, have been expanded by utilizing transferring cytotoxic T lymphocyte (CTL) (95) and transferring TCR gene (96). It is also necessary that they differentiate into double positive cells (CD4/8+), before generating iPSC-derived CTL (iCTL), by IL-2 which is the major cytokine in T-cell differentiation (97, 98). Experimentally tumor antigen-specific T lymphocytes can be generated in vitro by reprogramming designated T cells into iPSCs which are then differentiate back into T lymphocytes for infusion in patients. However, the safety of T cellderived human iPSCs must be more credible (99, 100).

Since most cancers in a specific tissue include acquired nucleotide mutations, iPSCs resulted from other normal tissues of the same patient hypothetically can be used to regenerate those injured tissues by the tumors themselves or subsequent (101,However, treatments 102). applications the iPSC-derived tissues in cancer therapy, needs that confiremed by in vivo studies. It has been made clear that human iPSCs derived from T lymphocytes maintain the pre-rearranged T cell receptor (TCR) gene; therefore, these iPSCs can be induced differentiation for functionally active T cells (93, 94). In 2018, Chao et al., described the generation of AML-iPSCs from two patients, with more emphasis on the notion that reprogramming of primary leukemias was not an unimportant assignment (103). Nevertheless, technical natural or reprogramming difficulties are significant barriers to reprogramming of primary

acute leukemic cells that need further investigation.

The differences in reprogramming prerearranged cells in AML and B-ALL propose that epigenetic reprogramming may be dependent on the specific genetic alterations and leukemia tumor subtype (104-106). Recent studies have shown that dendritic cells derived from human iPSCs can be completely functional for the assessment of hematopoietic diseases (107-109). However, these cells still need in vivo testing to confirm the effectiveness and safety of their application.

2- Induced NK Cells

Human natural killer (NK) cells are considered as an significant part of the innate immune system by producing significant cytokines for killing virally infected and/or malignant cells (110). While antitumor T-cell immune responses involve a diagnostic initiating phase and their responses are restricted to human leukocyte antigen (HLA), mature NK cells effectively kill malignant cells without any previous exposure (111). In addition, NK cells are the link between innate and adaptive immune systems that can increase and accelerate immunologic responses using their memory ability. At present, these cells are extracted by leukapheresis from the blood circulation and then separated from other cells via magnetic beads coated with anti-CD56. Both NK cell - and T cell -based adoptive immunotherapies have been used to treat patients with tumor malignancies. In recent studies, CD34⁺ hematopoietic progenitor cells derived from umbilical cord blood (UCB) are also being used as a basis for the generation of many numbers of allogeneic NK cells (112). Some research groups have defined various protocols for the production of NK cells from CD34⁺ cells using culture of stromal cell lines and a combination of cytokines that progress the expansion of NK cells (113-115). Furthermore, other researchers have been

able to produce large numbers of UCB CD34⁺ cells-derived NK-cells for adoptive immunotherapy in large-scale bioreactors, and for the use in future clinical cancer trials (116, 117). According to the studies on induced NK cells based on iPSC (iNK cells), reprogrammed NK cell can supply the desired volume of cell product to be used in leukemia therapy panel without any immunological response.

Conclusion

Induced pluripotent stem cells provide an alternative approach to human embryonic stem cells (hESCs) without any ethical concerns and with universal usage. Since they are obtained from dedifferentiation of adult cells and not from embryo, iPSCs overcome the ethical hurdles of embryonic stem cells. There are still significant obstacles that need to be considered. Despite the attractive research on iPSCderived cell transplantation in the grafts, there is still a potential risk of tumor formation. It is possible to generate patient specific iPSC from somatic tissues, which can contribute to drug development, disease modeling, and autologous stem cell therapy. Stem cells migrate to solid tumors and facilitate site-specific antitumor drug delivery. It is clear that induced hematopoietic stem cells -derived immune cells provide genetic manageable system to study human immune cell function and progress. In addition, these immune cells provide a main source of lymphocytes and may open new doors for cancer therapy. Finally, these studies have demonstrated that cell reprogramming in primary tumor cells is more difficult, and that further technological development is needed to be able to generate reliable iPSC models of cancers.

Conflicts of interest

There are no conflicts of interest.

References

- 1. Colman A. Induced pluripotent stem cells and human disease. Cell Stem Cell 2008;3(3):236-237.
- 2. Xia X, Chu J, Chen X. Induced pluripotent stem cells generated from reprogramming differentiated cells by defined factors. Sheng Wu Gong Cheng Xue Bao 2008;24(7):1121-1127.
- 3. Till JE, Mc CE. A direct measurement of the radiation sensitivity of normal mouse bone marrow cells. Radiat Res 1961;14:213-222.
- 4. Osawa M, Nakamura K, Nishi N, Takahasi N, Tokuomoto Y, Inoue H, et al. In vivo self-renewal of c-Kit+ Sca-1+Lin(low/-) hemopoietic stem cells. J Immunol 1996;156(9):3207-3214.
- 5. Spangrude GJ, Heimfeld S, Weissman IL. Purification and characterization of mouse hematopoietic stem cells. Science 1988;241(4861):58-62.
- 6. Okita K, Ichisaka T, Yamanaka S. Generation of germline-competent induced pluripotent stem cells. Nature 2007;448(7151):313-317.
- 7. Bindhya S, Sidhanth C, Shabna A, Krishnapriya S, M G, Ganesan TS. Induced Pluripotent Stem Cells: A New Strategy to Model Human Cancer. Int J Biochem Cell Biol 2019;107:62-68.
- 8. Camara DA, Mambelli LI, Porcacchia AS, Kerkis I. Advances and Challenges on Cancer Cells Reprogramming Using Induced Pluripotent Stem Cells Technologies. J Cancer 2016;7(15):2296-2303.
- 9. Chamberlain SJ, Li XJ, Lalande M. Induced pluripotent stem (iPS) cells as in vitro models of human neurogenetic disorders. Neurogenetics 2008;9(4):227-235.
- 10. Durcova-Hills G. Induced reprogramming of human somatic cells into pluripotency: a new way how to generate pluripotent stem cells. Differentiation 2008;76(4):323-325.
- 11. Aoi T. Advance in study of induced pluripotent stem cells (iPS cells). Nihon Rinsho 2008;66(5):850-856.

- 12. Beyene R, Boockvar JA. Disease-specific induced pluripotent stem cells. Neurosurgery 2008;63(6):12-18.
- 13. Dimos JT, Rodolfa KT, Niakan KK, Weisenthal LM, Mitsumoto H, Chung W, et al. Induced pluripotent stem cells generated from patients with ALS can be differentiated into motor neurons. Science 2008;321(5893):1218-1221.
- 14. Zhu D, Kong CSL, Gingold JA, Zhao R, Lee DF. Induced Pluripotent Stem Cells and Induced Pluripotent Cancer Cells in Cancer Disease Modeling. Adv Exp Med Biol 2018;1119:169-183
- 15. Yu J, Vodyanik MA, Smuga-Otto K, Antosiewicz-Bourget J, Frane JL, Tian S, et al. Induced pluripotent stem cell lines derived from human somatic cells. Science 2007;318(5858):1917-1920.
- 16. Duinsbergen D, Salvatori D, Eriksson M, Mikkers H. Tumors originating from induced pluripotent stem cells and methods for their prevention. Ann N Y Acad Sci 2009;1176:197-204.
- 17. Yuan TF, Arias-Carrion O. Locally induced neural stem cells/pluripotent stem cells for in vivo cell replacement therapy. Int Arch Med 2008;1(1):17-20.
- 18. Yaddanapudi K, Li C, Eaton JW. Vaccination with induced pluripotent stem cells confers protection against cancer. Stem Cell Investig 2018;5:23-28.
- 19. Tulloch NL, Pabon L, Murry CE. Get with the (re)program: cardiovascular potential of skin-derived induced pluripotent stem cells. Circulation 2008;118(5):472-475.
- 20. Stadtfeld M, Nagaya M, Utikal J, Weir G, Hochedlinger K. Induced pluripotent stem cells generated without viral integration. Science 2008;322(5903):945-949.
- 21. Shi Y, Do JT, Desponts C, Hahm HS, Scholer HR, Ding S. A combined chemical and genetic approach for the generation of induced pluripotent stem cells. Cell Stem Cell 2008;2(6):525-528.
- 22. Sato M, Kawana K, Adachi K, Fujimoto A, Yoshida M, Nakamura H, et al. Regeneration of cervical reserve cell-

- like cells from human induced pluripotent stem cells (iPSCs): A new approach to finding targets for cervical cancer stem cell treatment. Oncotarget 2017;8(25):40935-40945.
- 23. Park IH, Lerou PH, Zhao R, Huo H, Daley GQ. Generation of human-induced pluripotent stem cells. Nat Protoc 2008;3(7):1180-1186.
- 24. Singhal DK, Singhal R, Malik HN, Singh S, Kumar S, Kaushik JK, et al. Molecular cloning and production of caprine recombinant Oct4 protein for generation induced pluripotent stem cells. Mol Biol Rep 2015;42(12):1583-1591.
- 25. Blelloch R, Venere M, Yen J, Ramalho-Santos M. Generation of induced pluripotent stem cells in the absence of drug selection. Cell Stem Cell 2007;1(3):245-247.
- 26. Kim JB, Zaehres H, Wu G, Gentile L, Ko K, Sebastiano V, et al. Pluripotent stem cells induced from adult neural stem cells by reprogramming with two factors. Nature 2008;454(7204):646-650.
- 27. Nakagawa M, Koyanagi M, Tanabe K, Takahashi K, Ichisaka T, Aoi T, et al. Generation of induced pluripotent stem cells without Myc from mouse and human fibroblasts. Nat Biotechnol 2008;26(1):101-106.
- 28. Liu H, Zhu F, Yong J, Zhang P, Hou P, Li H, et al. Generation of induced pluripotent stem cells from adult rhesus monkey fibroblasts. Cell Stem Cell 2008;3(6):587-590.
- 29. Liao J, Cui C, Chen S, Ren J, Chen J, Gao Y, et al. Generation of induced pluripotent stem cell lines from adult rat cells. Cell Stem Cell 2009;4(1):11-15.
- 30. Singh VK, Kumar N, Kalsan M, Saini A, Chandra R. Mechanism of Induction: Induced Pluripotent Stem Cells (iPSCs). J Stem Cells 2015;10(1):43-62.
- 31. Park IH, Arora N, Huo H, Maherali N, Ahfeldt T, Shimamura A, et al. Disease-specific induced pluripotent stem cells. Cell 2008;134(5):877-886.
- 32. Nishikawa S, Goldstein RA, Nierras CR. The promise of human

- induced pluripotent stem cells for research and therapy. Nat Rev Mol Cell Biol 2008;9(9):725-729.
- 33. Zhang DM, Li JJ, Yan P, Hu JT. Establishment and identification of induced pluripotent stem cells in liver cancer patients. Asian Pac J Trop Med 2014;7(4):253-256.
- 34. Chen L, Kasai T, Li Y, Sugii Y, Jin G, Okada M, et al. A model of cancer stem cells derived from mouse induced pluripotent stem cells. PLoS One 2012;7(4):e33544-e33450.
- 35. Geoghegan E, Byrnes L. Mouse induced pluripotent stem cells. Int J Dev Biol 2008;52(8):1015-1022.
- 36. Aasen T, Raya A, Barrero MJ, Garreta E, Consiglio A, Gonzalez F, et al. Efficient and rapid generation of induced pluripotent stem cells from human keratinocytes. Nat Biotechnol 2008;26(11):1276-1284.
- 37. Anguera MC, Sadreyev R, Zhang Z, Szanto A, Payer B, Sheridan SD, et al. Molecular signatures of human induced pluripotent stem cells highlight sex differences and cancer genes. Cell Stem Cell 2012;11(1):75-90.
- 38. Eminli S, Utikal J, Arnold K, Jaenisch R, Hochedlinger K. Reprogramming of neural progenitor cells into induced pluripotent stem cells in the absence of exogenous Sox2 expression. Stem Cells 2008;26(10):2467-2474.
- 39. Yang J, Lam DH, Goh SS, Lee EX, Zhao Y, Tay FC, et al. Tumor tropism of intravenously injected human-induced pluripotent stem cell-derived neural stem cells and their gene therapy application in a metastatic breast cancer model. Stem Cells 2012;30(5):1021-1029.
- 40. Wartenberg M, Donmez F, Ling FC, Acker H, Hescheler J, Sauer H. Tumor-induced angiogenesis studied in confrontation cultures of multicellular tumor spheroids and embryoid bodies grown from pluripotent embryonic stem cells. FASEB J 2001;15(6):995-1005.

- 41. Chen L, Mizutani A, Kasai T, Yan T, Jin G, Vaidyanath A, et al. Mouse induced pluripotent stem cell microenvironment generates epithelial-mesenchymal transition in mouse Lewis lung cancer cells. Am J Cancer Res 2014;4(1):80-88.
- 42. Kim H, Schaniel C. Modeling Hematological Diseases and Cancer With Patient-Specific Induced Pluripotent Stem Cells. Front Immunol 2018;9:2243-2248.
- 43. Yang M, Liu Y, Hou W, Zhi X, Zhang C, Jiang X, et al. Mitomycin C-treated human-induced pluripotent stem cells as a safe delivery system of gold nanorods for targeted photothermal therapy of gastric cancer. Nanoscale 2017;9(1):334-340.
- 44. Crespo M, Vilar E, Tsai SY, Chang K, Amin S, Srinivasan T, et al. Corrigendum: Colonic organoids derived from human induced pluripotent stem cells for modeling colorectal cancer and drug testing. Nat Med 2018;24(4):526-531.
- 45. Fernandez Tde S, De Souza Fernandez C, Mencalha AL. Human induced pluripotent stem cells from basic research to potential clinical applications in cancer. Biomed Res Int 2013;2013:430290-430299.
- 46. Marin Navarro A, Susanto E, Falk A, Wilhelm M. Modeling cancer using patient-derived induced pluripotent stem cells to understand development of childhood malignancies. Cell Death Discov 2018;4:7-10.
- 47. Zielske SP, Spalding AC, Wicha MS, Lawrence TS. Ablation of breast cancer stem cells with radiation. Transl Oncol 2011;4(4):227-233.
- 48. Hu Y, Fu L. Targeting cancer stem cells: a new therapy to cure cancer patients. Am J Cancer Res 2012;2(3):340-356.
- 49. Hirschi KK, Li S, Roy K. Induced pluripotent stem cells for regenerative medicine. Annu Rev Biomed Eng 2014;16:277-294.
- 50. Saldanha SN, Royston KJ, Udayakumar N, Tollefsbol TO. Epigenetic

- Regulation of Epidermal Stem Cell Biomarkers and Their Role in Wound Healing. Int J Mol Sci 2015;17(1):e16-e20. 51. Luo Y, Liu C, Cerbini T, San H, Lin Y, Chen G, et al. Stable enhanced green fluorescent protein expression after differentiation and transplantation of reporter human induced pluripotent stem cells generated by AAVS1 transcription activator-like effector nucleases. Stem Cells Transl Med 2014;3(7):821-835.
- 52. Zhang CL, Huang T, Wu BL, He WX, Liu D. Stem cells in cancer therapy: opportunities and challenges. Oncotarget 2017;8(43):75756-75766.
- 53. Schriebl K, Satianegara G, Hwang A, Tan HL, Fong WJ, Yang HH, et al. Selective removal of undifferentiated human embryonic stem cells using magnetic activated cell sorting followed by a cytotoxic antibody. Tissue Eng Part A 2012;18(9-10):899-909.
- 54. Xu G, McLeod HL. Strategies for enzyme/prodrug cancer therapy. Clin Cancer Res 2001;7(11):3314-3324.
- 55. Malecki M, LaVanne C, Alhambra D, Dodivenaka C, Nagel S, Malecki R. Safeguarding Stem Cell-Based Regenerative Therapy against Iatrogenic Cancerogenesis: Transgenic Expression of DNASE1, DNASE1L3, DNASE2, DFFB Controlled By POLA1 Promoter in Proliferating and Directed Differentiation Resisting Human Autologous Pluripotent Induced Stem Cells Leads to their Death. J Stem Cell Res Ther 2013; 9(5): 21559-21567.
- 56. Chao MP, Gentles AJ, Chatterjee S, Lan F, Reinisch A, Corces MR, et al. Human AML-iPSCs Reacquire Leukemic Properties after Differentiation and Model Clonal Variation of Disease. Cell Stem Cell 2017;20(3):329-344.
- 57. Griscelli F, Oudrhiri N, Feraud O, Divers D, Portier L, Turhan AG, et al. Generation of induced pluripotent stem cell (iPSC) line from a patient with triple negative breast cancer with hereditary exon 17 deletion of BRCA1 gene. Stem Cell Res 2017;24:135-138.

- 58. Zeng S, Liu L, Ouyang Q, Zhao Y, Lin G, Hu L, et al. Generation of induced pluripotent stem cells (iPSCs) from a retinoblastoma patient carrying a c.2663G>A mutation in RB1 gene. Stem Cell Res 2016;17(2):208-211.
- 59. Liu H, Kim Y, Sharkis S, Marchionni L, Jang YY. In vivo liver regeneration potential of human induced pluripotent stem cells from diverse origins. Sci Transl Med 2011;3(82):82-93.
- 60. Knorr DA, Ni Z, Hermanson D, Hexum MK, Bendzick L, Cooper LJ, et al. Clinical-scale derivation of natural killer cells from human pluripotent stem cells for cancer therapy. Stem Cells Transl Med 2013;2(4):274-283.
- 61. Vizcardo R, Masuda K, Yamada D, Ikawa T, Shimizu K, Fujii S, et al. Regeneration of human tumor antigenspecific T cells from iPSCs derived from mature CD8(+) T cells. Cell Stem Cell 2013;12(1):31-36.
- 62. Gagne AL, Maguire JA, Gandre-Babbe S, Chou ST, Tasian SK, Loh ML, et al. Generation of a human Juvenile myelomonocytic leukemia iPSC line, CHOPi001-A, with a mutation in CBL. Stem Cell Res 2018;31:157-160.
- 63. Caren H, Stricker SH, Bulstrode H, Gagrica S, Johnstone E, Bartlett TE, et al. Glioblastoma Stem Cells Respond to Differentiation Cues but Fail to Undergo Commitment and Terminal Cell-Cycle Arrest. Stem Cell Reports 2015;5(5):829-842.
- 64. Hu K, Yu J, Suknuntha K, Tian S, Montgomery K, Choi KD, et al. Efficient generation of transgene-free induced pluripotent stem cells from normal and neoplastic bone marrow and cord blood mononuclear cells. Blood 2011;117(14):e109-119.
- 65. Kim J, Hoffman JP, Alpaugh RK, Rhim AD, Reichert M, Stanger BZ, et al. An iPSC line from human pancreatic ductal adenocarcinoma undergoes early to invasive stages of pancreatic cancer progression. Cell Rep 2013;3(6):2088-2099.

- 66. Aponte PM, Caicedo A. Stemness in Cancer: Stem Cells, Cancer Stem Cells, and Their Microenvironment. Stem Cells Int 2017;2017;5619472-5619479.
- 67. Van Zant G, Liang Y. Concise review: hematopoietic stem cell aging, life span, and transplantation. Stem Cells Transl Med 2012;1(9):651-657.
- 68. Bryder D, Rossi DJ, Weissman IL. Hematopoietic stem cells: the paradigmatic tissue-specific stem cell. Am J Pathol 2006;169(2):338-346.
- 69. Singh VK, Kalsan M, Kumar N, Saini A, Chandra R. Induced pluripotent stem cells: applications in regenerative medicine, disease modeling, and drug discovery. Front Cell Dev Biol 2015;3:2-9.
- 70. Sankar S, Sharma CS, Rath SN, Ramakrishna S. Electrospun Fibers for Recruitment and Differentiation of Stem Cells in Regenerative Medicine. Biotechnol J 2017;12(12): 1700263-1700269.
- 71. Harding J, Mirochnitchenko O. Preclinical studies for induced pluripotent stem cell-based therapeutics. J Biol Chem 2014;289(8):4585-4593.
- 72. Kondo Y, Toyoda T, Inagaki N, Osafune K. iPSC technology-based regenerative therapy for diabetes. J Diabetes Investig 2018;9(2):234-243.
- 73. Patel M, Yang S. Advances in reprogramming somatic cells to induced pluripotent stem cells. Stem Cell Rev 2010;6(3):367-380.
- 74. Zuo Y, Su G, Cheng L, Liu K, Feng Y, Wei Z, et al. Coexpression analysis identifies nuclear reprogramming barriers of somatic cell nuclear transfer embryos. Oncotarget 2017;8(39):65847-65859.
- 75. Tian XC, Kubota C, Enright B, Yang X. Cloning animals by somatic cell nuclear transfer--biological factors. Reprod Biol Endocrinol 2003;1:98-100.
- 76. Mall M, Wernig M. The novel tool of cell reprogramming for applications in molecular medicine. J Mol Med (Berl) 2017;95(7):695-703.

- 77. Mayhew CN, Wells JM. Converting human pluripotent stem cells into beta-cells: recent advances and future challenges. Curr Opin Organ Transplant 2010;15(1):54-60.
- 78. Kawamata M, Suzuki A. Cell fate modification toward the hepatic lineage by extrinsic factors. J Biochem 2017;162(1):11-16.
- 79. Renga M, Pedrazzoli P, Siena S. Present results and perspectives of allogeneic non-myeloablative hematopoietic stem cell transplantation for treatment of human solid tumors. Ann Oncol 2003;14(8):1177-1184.
- 80. Karadurmus N, Sahin U, Bahadir Basgoz B, Arpaci F, Demirer T. A Review of Allogeneic Hematopoietic Stem Cell Transplantation in Metastatic Breast Cancer. Int J Hematol Oncol Stem Cell Res 2018;12(2):111-116.
- 81. Hatzimichael E, Tuthill M. Hematopoietic stem cell transplantation. Stem Cells Cloning 2010;3:105-117.
- 82. Birbrair A, Frenette PS. Niche heterogeneity in the bone marrow. Ann N Y Acad Sci 2016;1370(1):82-96.
- 83. Maali A, Atashi A, Ghaffari S, Kouchaki R, Abdolmaleki F, Azad M. A Review on Leukemia and iPSC Technology: Application in Novel Treatment and Future. Curr Stem Cell Res Ther 2018;13(8):665-675.
- 84. Szabo E, Rampalli S, Risueno RM, Schnerch A, Mitchell R, Fiebig-Comyn A, et al. Direct conversion of human fibroblasts to multilineage blood progenitors. Nature 2010;468(7323):521-526.
- 85. Doulatov S, Vo LT, Chou SS, Kim PG, Arora N, Li H, et al. Induction of multipotential hematopoietic progenitors from human pluripotent stem cells via respecification of lineage-restricted precursors. Cell Stem Cell 2013;13(4):459-470.
- 86. Perez-Pinera P, Ousterout DG, Gersbach CA. Advances in targeted genome editing. Curr Opin Chem Biol 2012;16(3-4):268-277.

- 87. Hotta A, Yamanaka S. From Genomics to Gene Therapy: Induced Pluripotent Stem Cells Meet Genome Editing. Annu Rev Genet 2015;49:47-70.
- 88. Larson S, De Oliveira SN. Genemodified hematopoietic stem cells for cancer immunotherapy. Hum Vaccin Immunother 2014;10(4):982-985.
- 89. Gschweng E, De Oliveira S, Kohn DB. Hematopoietic stem cells for cancer immunotherapy. Immunol Rev 2014;257(1):237-249.
- 90. Barrett DM, Grupp SA, June CH. Chimeric Antigen Receptor- and TCR-Modified T Cells Enter Main Street and Wall Street. J Immunol 2015;195(3):755-761.
- 91. Jiang Z, Han Y, Cao X. Induced pluripotent stem cell (iPSCs) and their application in immunotherapy. Cell Mol Immunol 2014;11(1):17-24.
- 92. Jiang Y, Habibollah S, Tilgner K, Collin J, Barta T, Al-Aama JY, et al. An induced pluripotent stem cell model of hypoplastic left heart syndrome (HLHS) reveals multiple expression and functional differences in HLHS-derived cardiac myocytes. Stem Cells Transl Med 2014;3(4):416-423.
- 93. Vizcardo R, Klemen ND, Islam SMR, Gurusamy D, Tamaoki N, Yamada D, et al. Generation of Tumor Antigen-Specific iPSC-Derived Thymic Emigrants Using a 3D Thymic Culture System. Cell Rep 2018;22(12):3175-3190.
- 94. Saito H, Okita K, Chang AE, Ito F. Adoptive Transfer of CD8+ T Cells Generated from Induced Pluripotent Stem Cells Triggers Regressions of Large Tumors Along with Immunological Memory. Cancer Res 2016;76(12):3473-3483.
- 95. Chapuis AG, Ragnarsson GB, Nguyen HN, Chaney CN, Pufnock JS, Schmitt TM, et al. Transferred WT1-reactive CD8+ T cells can mediate antileukemic activity and persist in post-transplant patients. Sci Transl Med 2013;5(174):174-179.

- 96. Robbins PF, Morgan RA, Feldman SA, Yang JC, Sherry RM, Dudley ME, et al. Tumor regression in patients with metastatic synovial cell sarcoma and melanoma using genetically engineered lymphocytes reactive with NY-ESO-1. J Clin Oncol 2011;29(7):917-924.
- 97. Maeda T, Nagano S, Ichise H, Kataoka K, Yamada D, Ogawa S, et al. Regeneration of CD8alphabeta T Cells from T-cell-Derived iPSC Imparts Potent Tumor Antigen-Specific Cytotoxicity. Cancer Res 2016;76(23):6839-6850.
- 98. Hoyer S, Prommersberger S, Pfeiffer IA, Schuler-Thurner B, Schuler G, Dorrie J, et al. Concurrent interaction of DCs with CD4(+) and CD8(+) T cells improves secondary CTL expansion: It takes three to tango. Eur J Immunol 2014;44(12):3543-3559.
- 99. Kaneko S. In Vitro Generation of Antigen-Specific T Cells from Induced Pluripotent Stem Cells of Antigen-Specific T Cell Origin. Methods Mol Biol 2016;1393:67-73.
- 100. Brauer PM, Singh J, Xhiku S, Zuniga-Pflucker JC. T Cell Genesis: In Vitro Veritas Est? Trends Immunol 2016;37(12):889-901.
- 101. Papapetrou EP. Patient-derived induced pluripotent stem cells in cancer research and precision oncology. Nat Med 2016;22(12):1392-1401.
- 102. Mahla RS. Stem Cells Applications in Regenerative Medicine and Disease Therapeutics. Int J Cell Biol 2016;2016:6940283-6940288.
- 103. Chao HM, Chern E. Patient-derived induced pluripotent stem cells for models of cancer and cancer stem cell research. J Formos Med Assoc 2018;117(12):1046-1057.
- 104. Munoz-Lopez A, Romero-Moya D, Prieto C, Ramos-Mejia V, Agraz-Doblas A, Varela I, et al. Development Refractoriness of MLL-Rearranged Human B Cell Acute Leukemias to Reprogramming into Pluripotency. Stem Cell Reports 2016;7(4):602-618.

- 105. McClellan JS, Dove C, Gentles AJ, Ryan CE, Majeti R. Reprogramming of primary human Philadelphia chromosome-positive B cell acute lymphoblastic leukemia cells into nonleukemic macrophages. Proc Natl Acad Sci U S A 2015;112(13):4074-4079.
- 106. Wouters BJ, Delwel R. Epigenetics and approaches to targeted epigenetic therapy in acute myeloid leukemia. Blood 2016;127(1):42-52.
- 107. Li Y, Liu M, Yang ST. Dendritic cells derived from pluripotent stem cells: Potential of large scale production. World J Stem Cells 2014;6(1):1-10.
- 108. Vo LT, Daley GQ. De novo generation of HSCs from somatic and pluripotent stem cell sources. Blood 2015;125(17):2641-2648.
- 109. Iizuka-Koga M, Asashima H, Ando M, Lai CY, Mochizuki S, Nakanishi M, et al. Functional Analysis of Dendritic Cells Generated from T-iPSCs from CD4+ T Cell Clones of Sjogren's Syndrome. Stem Cell Reports 2017;8(5):1155-1163.
- 110. Paul S, Lal G. The Molecular Mechanism of Natural Killer Cells Function and Its Importance in Cancer Immunotherapy. Front Immunol 2017;8:1124-1128.
- 111. Sharma P, Kumar P, Sharma R. Natural Killer Cells Their Role in Tumour Immunosurveillance. J Clin Diagn Res 2017;11(8):BE01-BE05.
- 112. Spanholtz J, Preijers F, Tordoir M, Trilsbeek C, Paardekooper J, Witte T, et al. Clinical-grade generation of active NK cells from cord blood hematopoietic progenitor cells for immunotherapy using a closed-system culture process. PLoS One 2011;6(6):e20740-20745.
- 113. Luevano M, Madrigal A, Saudemont A. Generation of natural killer cells from hematopoietic stem cells in vitro for immunotherapy. Cell Mol Immunol 2012;9(4):310-320.
- 114. Cany J, Van der Waart AB, Spanholtz J, Tordoir M, Jansen JH, Van der Voort R, et al. Combined IL-15 and IL-12 drives the generation of CD34(+)-

derived natural killer cells with superior maturation and alloreactivity potential following adoptive transfer. Oncoimmunology 2015;4(7):e1017701-e1017705.

- 115. Dezell SA, Ahn YO, Spanholtz J, Wang H, Weeres M, Jackson S, et al. Natural killer cell differentiation from hematopoietic stem cells: a comparative analysis of heparin- and stromal cell-supported methods. Biol Blood Marrow Transplant 2012;18(4):536-545.
- 116. Eguizabal C, Zenarruzabeitia O, Monge J, Santos S, Vesga MA, Maruri N, et al. Natural killer cells for cancer immunotherapy: pluripotent stem cellsderived NK cells as an immunotherapeutic perspective. Front Immunol 2014;5:439-449.
- 117. Spanholtz J, Tordoir M, Eissens D, Preijers F, Van der Meer A, Joosten I, et al. High log-scale expansion of functional human natural killer cells from umbilical cord blood CD34-positive cells for adoptive cancer immunotherapy. PLoS One 2010;5(2):e9221-e9224.