Evaluation of Twenty Four Discriminant Indices for Differentiating Beta-Thalassemia Trait from Iron Deficiency Anemia in Egyptians

Shaimaa Abdelmalik Pessar, MD

Faculty of Medicine, Ain Shams University - Cairo, Egypt *Corresponding author: Shaimaa Abdelmalik Pessar, Clinical Pathology Department, Faculty of Medicine, Ain Shams University, Abbasseya, Cairo, Egypt, postal code: 11591. Email: <u>shaimaamalik@med.asu.edu.eg</u>. Orchid ID: <u>orchid.org/0000-0002-8947-2324</u>

Received: 10 February 2019 Accepted: 10 June 2019

Abstract

Background: Many Red Blood Cell (RBC) indices have been developed based on mathematical formulae to discriminate beta-thalassemia trait (β TT) from iron deficiency anemia (IDA). The latter two conditions represent the most common causes of microcytic hypochromic anemia in Egypt. This study aimed To evaluate the diagnostic reliability of 24 published discriminant indices for differentiating β TT from IDA in Egyptians.

Materials and Methods: A cross sectional study included a total of 166 subjects (108 IDA & 58 β TT) aged 1-18 years were recruited from Hematology laboratory of Pediatric Hospital, Ain Shams University, Cairo, Egypt. A full diagnostic algorithm was performed using complete blood count, hemoglobin electrophoresis by High Performance Liquid Chromatography (HPLC), iron profile and PCR detection of 22 mutations common for β TT. Twenty-four formulas were applied and their performance characteristics were calculated for each index.

Results: The highest accuracy (True positive + True negative/ All cases) & Youden's Index (Sensitivity+Specificity-100) were for Red Cell Distribution Width (RDWI) and Hameed index closely followed by Keikhaei index while the least performance was for RDW-SD, RDW-CV and Shine & Lal indices.

Conclusion The superiority of an index over another in distinguishing β TT from IDA allowed only partially better selection of cases warranting further confirmatory molecular studies. None of the studied formulae provided a surrogate test for Hb electrophoresis as mass screening.

Key words: Anemia, Beta-Thalassemia, Erythrocyte Indices, Iron deficiency.

Introduction

The two most frequent causes of microcytic anemia worldwide are beta thalassemia trait (BTT) and iron deficiency anemia (IDA) (1). In Egypt, both conditions represent a public health problem. About 64% of Egyptian children, especially from rural areas, have IDA (2). Also, the carrier rate of BTT ranges from 9-16.4% with an overall rate of 12.36% (3). Diagnosis of βTT often becomes perplexing due to its overlapping and similar features with IDA (4). Definitive methods for differential diagnosis between and IDA include quantitative βTT detection of HbA2 and DNA mutational analysis. While being accurate, these tests are too expensive and time-consuming for initial mass screening (5). Since screening will continue to be the cornerstone of the

strategies aimed at β -thalassemia control, attempts develop effective to and economic techniques for thalassemia screening have become essential in the countries with high prevalence of such diseases (6). Automated cell counters could provide a rapid, technically reliable method for effective population screening of β TT (7). Red cell indices have been used in a number of formulae, designed to discriminate cases of iron deficiency from cases of βTT , each has some degree of inaccuracy (8-25). A test is diagnostically valuable if it shows: superior sensitivity, specificity, efficiency, Youden's index (which is highly sensitive to the spectrum of the disease in a studied population), high positive predictive value, negative predictive value, positive likelihood ratio, and low negative likelihood ratio (26).

Measures of diagnostic accuracy are very sensitive to the characteristics of the population in which the test is evaluated. It is therefore of utmost importance to know how to interpret them as well as when and under what conditions to use them (27). This study intended to evaluate the diagnostic reliability of 24 published Differentiating Formulas (DFs) in detecting Egyptian children and adolescents who have BTT.

Materials and Methods

This cross sectional study was carried out on 280 microcytic hypochromic children and adolescent recruited from a total population of 1627 (17.2%)that presented the to outpatient clinics of Pediatric Hospital, Ain Shams University, Cairo, Egypt. Out of the 280 microcytic hypochromic children and adolescents, a total of 166 children and adolescents (108 IDA & 58 β TT) aged 1-18 years were enrolled in the study after procuring informed consents from their care givers. Out of the 166, 103 (62%) were males and 63 (38%) were females with a male to female ratio of 1.6:1. They were then divided into 108 IDA and 58 BTT according to the results of their iron profile, Hb A2 quantitation by High Performance Liquid Chromatography (HPLC), and beta thalassemia gene mutation testing by Polymerase Chain Reaction (PCR).

All subjects underwent the following procedures:

- Complete blood count (Coulter Gen System 2, Beckman-Coulter, Miami, FL, USA).
- Hb A₂ quantitation (βTT short dual program, D10, Bio-Rad Laboratories, California, USA), D10 is a compact high performance liquid chromatograph designed for simple and fully automated measurement of HbA1c, Ao, F and for the detection of abnormal Hbs.
- Serum iron and total iron binding capacity measure (Colorimetrically by Olympus

AU 600 autoanalyser, Olympus Diagnostics GmbH, Hamburg, Germany), and_serum ferritin (radioimmunoassay by ARCHITECT, Abbott Diagnostics, Wiesbaden, Germany).

Detection of β -globin gene mutations (Reverse hybridization PCR, β-Globin Strip—Assay MED, Vienna Lab Diagnostics, GmbH). The **B**-Globin StripAssay MED[™] is based on the reverse-hybridization principle, and includes three successive steps: DNA is isolated from samples by a rapid and convenient procedure then β -globin gene sequences are in vitro amplified and biotin-labeled in a single (multiplex) amplification Finally, reaction. the amplification products are selectively hybridized to a test strip, which contains oligonucleotide probes (wild type- and mutant-specific) immobilized as parallel lines. Bound biotinylated sequences are detected using streptavidin-alkaline phosphatase and color substrates. The assay covers 22 mutations characteristic for Mediterranean area: $-101[C \rightarrow T]$, -87 $[C \rightarrow G]$, -30 $[T \rightarrow A]$, codon 5 [-CT], codo 6 [G \rightarrow A] HbC, codon 6 [A \rightarrow T] HbS, codon 6 [-A], codon 8 [-AA], codon 8/9 [+G], codon 15 [TGG \rightarrow TGA], codon 27 $[G \rightarrow T]$ (knossos), IVS I-1 $[G \rightarrow A]$, IVS I-5 [G \rightarrow C], IVS I-6 [T \rightarrow C], IVS I-110 $[G \rightarrow A]$, IVS I-116 $[T \rightarrow G]$, IVS I-130 $[G \rightarrow C]$, codon 39 $[C \rightarrow T]$, codon 44 [-C], IVS II-1 [G \rightarrow A], IVS II-745 [C \rightarrow G], IVS II-848 [C \rightarrow A]. The study was conducted in accordance with the stipulations of the local ethical and scientific committees of Ain Shams University; Egypt and the procedures respected the ethical standards in Helsinki declaration of 1964. The 24 formulas were calculated for all cases. (Table I) (8-25).

Statistical analysis

Quantitative variables were presented either as mean and standard deviation (SD) or median and range, and qualitative variables as number and percentage. Quantitative variables were compared using unpaired t-test; Analysis of data was done on an IBM computer using the Statistical Package for the Social Sciences, version 12 (SPSS Inc., Chicago, IL, USA) and were evaluated at the 0.05 significance level. The diagnostic value of each formula was performed by calculating the Sensitivity, Specificity, Positive Predictive Value (PPV), Negative Predictive Value (NPV), Positive likelihood Ratio (LR+), Negative likelihood Ratio (LR-), Diagnostic Accuracy (efficiency); True positive + True negative/ All cases & Youden Index (Sensitivity+Specificity-100).

Results

Comparison of routine hematologic parameters are shown in Table II. Genetic mutations in β TT subjects: The four most common mutations were, in order of decreasing frequency, IVS-1-6 (T>C) (16/58, 27.6%), IVS-I-110 (G>A) (13/58,

22.4%), IVS-I-1 (G>A) (12/58, 20.7) and IVS-II-745 (C>G) (7/58, 12.1%) followed by other less frequent mutations such as IVS-II-848 (C>A) and codon 5 (-CT) (3/58, 5.2% for each), then IVS-II-1 (G>A) (2/58, 3.4%), and at last, -87 (C>G), and codon 39 (C4T) (1/58, 1.7% for each) (Figures 1 and 2). Diagnostic Performance of 24 calculated RBCsindices-based formulae: The diagnostic performance of each formula is represented by its efficiency (diagnostic accuracy). Our study revealed that the highest accuracy (efficiency) and Youden index were for Red Cell Distribution Width (RDWI), Hameed index and Keikhaei index with the least performance and Youden index were for RDW-SD, Shine & Lal and Bessman & Feinstein indices (RDW-CV) (Table III and Figure 3).

Index	Formula	βΤΤ	IDA
Mentzer index (MI)	MCV/RBCs count	<13	>13
Modified Mentzer index		<12	>14
England and Fraser index (E&FI)	MCV-RBCs-(5xHb)-K K is calculated to be 8.4	<0	>0
Shine and Lal index (S&LI)	MCV ² xMCHx0.01	<1530	>1530
Green and King index (G&KI)	MCV ² xRDW/100xHb	<65	>65
Srivastava and Bevington index (S&BI)	MCH/RBCs	<3.8	>3.8
Red cell distribution width index (RDWI)	MCVxRDW/RBCs	<220	>220
Ricerca et al index (RI)	RDW/RBCs	<3.3	>3.3
Red Blood Cell Count (RBCC)	RBCs count	>5	<5
Bessman and Feinstein index (B&FI)	RDW-CV	<14	>14
RDW-SD	RDW-SD	<46	>46
Keikhaei index (KI)	Hb×RDW×100/ (RBC)2×MCHC	<21	>21
Ehsani et al index (EI)	MCV-10×RBC	<15	>15
Sirdah et al index (SI)	MCV-RBC-3×Hb	<27	>27
Mean Cell Hemoglobin Density (MCHD)	MCH/MCV	> 0.3045	< 0.3045
Telmissani et al index (MDHL)	MCHD×RBC	>1.7M, >1.5F	<1.7M, <1.5F
Sirachainan et al	1.5 Hb-0.05 MCV	>14	<14
Bordbar et al	80-MCV × 27-MCH	>44.76	<44.76
Differentiating score (DS)	Male: (0.096xMCV)+(0.415xRDW)-(0.139xRBCs)-12.722 Female: (0.096xMCV)+(0.415xRDW)-12.722	<0.3095	>0.3095
Huber-Herklotz Index (HH)	(MCH × RDW × 0.1/RBC)+ RDW	<21	>23
Kerman index 1	MCVxMCH/RBCs	<250	321-370
Kerman index 2	MCVxMCH/RBCsxMCHC	<8	10.5-13
Hisham index (Hi)	(MCH×RDW)/RBC	< 67	≥67
Hameed index (Ha)	(MCH×Hct×RDW)/ (RBC×Hb) ²	< 220	≥220

Table (I): the discriminative indices with their respective cut-off values:

Iran J Ped Hematol Oncol. 2019, Vol 9. No 3, 135-146

Variables	βTT (n=58)	IDA (n=108)	Р
RBCs (x10 ⁶ /uL)	5.4 <u>+</u> 0.7	4.5 <u>+</u> 0.5	0.02 S
Hb (g/ dL)	10.6 <u>+</u> 1.5	9.4 <u>+</u> 1.8	0.03 S
HCT (%)	34 <u>+</u> 4.5	31 <u>+</u> 4	0.09 NS
MCV (fL)	63.9 <u>+</u> 4.5	65 <u>+</u> 6	<0.001 HS
MCH (pg)	19±1.8	19.7 <u>+</u> 3	0.06 NS
MCHC (g/dL)	31 <u>+</u> 1.7	30 <u>+</u> 2.7	0.1 NS
RDW (%)	17 <u>+</u> 3.4	19.5 <u>+</u> 4	<0.001 HS
HbA ₂ (%)	5.4 <u>+</u> 0.7	2.8 <u>+</u> 0.5	<0.001 HS
Serum iron (µg/dL)	39(28-60)	29(21-41)	0.07 NS
TIBC (µg/dL)	405 <u>+</u> 68	463 <u>+</u> 62	0.1 NS
Serum ferritin (ng/mL)	56(42-69)	12(7-20)	<0.001 HS

Table (II): Comparison of hematological parameters between both disorders:

βTT: beta thalassemia trait, IDA: iron deficiency anemia, S: significant, NS: non significant, HS: highly significant, RBCs: red blood cells, Hb: hemoglobin, Hct: hematocrit, MCV: mean corpuscular volume, MCH: mean corpuscular hemoglobin, MCHC: mean corpuscular hemoglobin concentration, RDW: red cell distribution width, TIBC: total iron binding capacity

Table (III): Performance criteria (sensitivity, specificity, PPV, NPV, LR+, LR-, accuracy and Youden index) for each discriminating index.

		Sensitivity	ndex) for eac Specificity	PPV	NPV	+ve LR	-ve LR	Accuracy	YI
MI	βΤΤ	68.96	63.88	50.63	88.46	1.9	0.48	65.66	32.84
	IDA	63.88	68.96	88.46	50.63	2.06	0.52		
Modified MI*	βττ	78.94	68.57	57.69	85.71	2.5	0.32	46.98	47.51
	IDA	68.57	78.94	85.71	57.69	3.25	0.4		
E&F Index	βΤΤ	63.79	74.07	56.92	79.21	2.46	0.49	70.48	37.86
Lai macx	IDA	74.07	63.79	79.21	56.92	2.04	0.41		
S&L Index	βΤΤ	100	0	34.94	0	1	0	34.94	0
JOE MILLER	IDA	0	100	0	34.94	0	1		
G&K Index	βΤΤ	50	89.81	72.5	76.98	4.90	0.56	75.90	39.81
Garcinaex	IDA	89.81	50	76.98	72.5	1.8	0.2		55.01
S&B Index	βΤΤ	55.17	66.66	47.05	37.47	1.65	0.67	62.65	21.83
Sub muck	IDA	66.66	55.17	37.47	47.05	1.49	0.6		
RDWI	βΤΤ	70.69	83.33	69.49	84.11	4.24	0.35	78.91	54.02
NDW1	IDA	83.33	70.69	84.11	69.49	2.84	0.23		
RI	βΤΤ	65.51	82.41	66.66	81.65	3.72	0.42	76.50	47.92
	IDA	82.41	65.51	81.65	66.66	1.0	0.27		
RBC count	βΤΤ	67.24	71.29	55.71	80.21	2.34	0.46	69.88	38.53
	IDA	71.29	67.24	80.21	55.71	2.17	0.43		
B&F Index	βττ	5.17	96.29	42.86	65.41	1.39	0.98	64.46	1.46
	IDA	96.29	5.17	65.41	42.86	1.02	0.72		
RDW-SD	βττ	10.34	87.04	30	46.38	0.8	1.03	60.24	-2.62
	IDA	87.04	10.34	46.38	30	0.97	1.25		
KI	βττ	58.62	89.81	75.55	80.16	5.75	0.46	78.91	48.43
	IDA	89.81	58.62	80.16	75.55	2.17	0.17		
EI	βττ	65.52	63.89	49.35	77.53	1.81	0.54	64.46	29.41
	IDA	63.89	65.52	77.53	49.35	1.85	0.55		
SI	βττ	46.55	84.26	61.36	74.59	2.96	0.63	71.08	30.81
	IDA	84.26	46.55	74.59	61.36	1.58	0.34		
MCHD	βττ	74.14	49.07	43.88	77.94	1.46	0.53	57.83	23.21
	IDA	49.07	74.14	77.94	43.88	1.90	0.69		
MDHL	βττ	62.07	83.33	66.67	80.36	3.72	0.46	75.90	45.4
	IDA	83.33	62.07	80.36	66.67	2.20	0.27		
Sirachainan et	βΤΤ	24.14	89.81	56	68.79	2.37	0.84	66.87	13.95
al	IDA	89.81	24.14	68.79	56	1.18	0.42		
Bordbar et al	βττ	94.83	24.07	40.14	89.66	1.25	0.21	48.79	18.9

S.A.

	IDA	24.07	94.83	89.66	40.14	4.65	0.8		
DS	βΤΤ	68.96	72.22	57.14	81.25	2.48	0.43	71.08	41.18
	IDA	72.22	68.96	81.25	57.14	2.33	0.4		
HH Index**	βΤΤ	32.43	95.79	75	78.45	7.7	0.7	62.05	28.22
	IDA	95.79	32.43	78.45	75	1.42	0.13		
Kerman index	βΤΤ	68	36.67	47.22	57.89	1.07	0.87	33.73	4.67
1***	IDA	36.67	68	57.89	47.22	1.15	0.93		
Kerman index	βΤΤ	80.49	42.62	48.53	76.47	1.40	0.46	35.54	23.11
2****	IDA	42.62	80.49	76.47	48.53	2.18	0.71		
Hisham index	βΤΤ	67.24	81.48	66.10	82.24	3.63	0.40	76.50	48.72
(Hi)	IDA	81.48	67.24	82.24	66.10	2.49	0.27		
Hameed index	βΤΤ	70.69	83.33	69.49	84.11	4.24	0.35	78.91	54.02
(Ha)	IDA	83.33	70.69	84.11	69.49	2.84	0.24		

*MMI had 58 indeterminate cases (12-14).** H-H index had 34 indeterminate cases (21-23). ***Kerman 1 had 22 indeterminate cases (250-320).****Kerman 2 had 26 indeterminate cases (8-10.4).

MI: Mentzer index, E&F: England and Fraser index, S&L: Shine and Lal index, G&K: Green and King index, S&B: Srivastava and Bevington index, RDWI: Red Cell Distribution Width index, RI: Ricerca index, B&F: Bessman and Feinstein index, RDW-SD: Red Cell Distribution Width-standard deviation, KI: Keikhaei index, EI: Ehsani index, SI: Sirdah index, MCHD: Mean Cell Hemoglobin Density, MDHL: Mean Density Of Hemoglobin Per Liter Of Blood or Telmissani et al index, DS: Differentiating score, HH: Huber-Herklotz index, βTT: beta thalassemia trait, IDA: iron deficiency anemia, PPV: Positive Predictive value, NPV: Negative Predictive value, LR+: Positive Likelihood ratio, LR-: Negative Likelihood ratio, YI: Youden index.

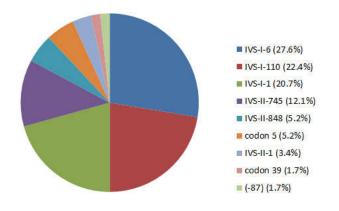
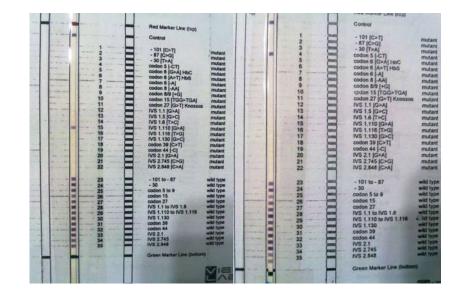


Figure 1: β thalassemia mutations frequency in carriers



Iran J Ped Hematol Oncol. 2019, Vol 9. No 3, 135-146

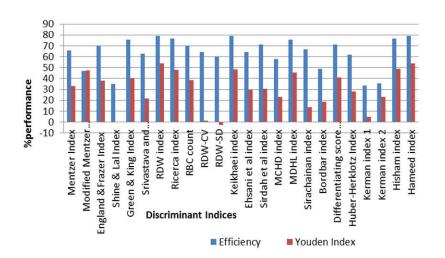


Figure 2: Lt: heterozygousity for IVS 1.110, Rt: heterozygosity for codon 5

Figure 3: Diagnostic Performance of The Studied Discriminant Indices

Discussion

The high carrier rate of β -thalassemia in Egypt is confounded by the phenotypic and genotypic heterogeneity attributed to its position as a liaison between the middle east and the Mediterranean basin. Given the high prevalence of each of IDA and β TT in the population; simple, costeffective and reliable screening tools for differentiation between both disorders are needed to avoid the use of the more laborious and expensive genetic studies. Many studies have evaluated the efficiency and discriminative power of various RBC indices and discriminating formulas which have come into interest. Individual hematological parameters as; Hb, RBCs count, Hb A2 and ferritin were significantly higher in βTT than IDA. Meanwhile, MCV and RDW were significantly lower in BTT than IDA. Reduction of MCV and MCH values in β TT did not correlate with the degree of anemia, similar findings were reported in Nesa et al (28) and Rathod et al (29). In this study, clinical, laboratory and genetic diagnoses were established for all the 166 cases; 108 IDA and 58 BTT. Then, the twenty four indices were retrospectively calculated for all cases and each index was evaluated for its discriminating power between IDA and BTT according to sensitivity, specificity, PPV, NPV, LR+ve, LR-ve, accuracy and Youden index. The reliability of various formulae studied in this work gave performance characteristics unique to their respective populations. In Kurdistan/Iraq, Hameed index and Hisham index achieved the highest youden indices followed by RDWI and Keikhaei index with the least reliable being S&L index. (25). High reliability of RDWI was reported by Nesa et al. (28) in Bangladesh and Sirdah et al. (18) in the Palestinian population together with their formula and G&K index. Alfadhli et al. (30) in Kuwait concluded that the E&F index was the best discriminant while the S&L index was found ineffective in the differentiation between IDA and β TT cases. Urrechaga (31) in Spain and Ntaios et al. (32) in Greece found that G&K index ranked the best with a Youden index of 80.9% and 70.86% respectively. In Pakistan, Niazi et al. (7) concluded that the accuracy and Youden index were highest for RDWI followed by Mentzer and S&L index while, Ehsani and colleagues (17) showed Mentzer as the best discrimination index followed by their new index in Pakistani patients. In India, Dhara and Harsh (33) introduced the RDW-SD as the best for distinguishing between IDA and BTT. They also considered both the RBC count and E&F index to be useful functions for that purpose while the RDW-CV to be ineffective whereas, Adlekha, et al. (4) found that RDWI and DS were of higher sensitivity and specificity than MDHL and MI in predicting BTT. In Turkey, similar results regarding the high reliability of RDWI were reported by Demir et al. (34) together with RBC count. Also, Beyan et al. (35) who concluded that none of the discriminant formulations was superior to RBC Count. While, Vehapoglu et al. (1) found that the highest efficiency was for the Mentzer index (91%) followed by the Ehsani et al. index (84.8%) then RBC count (83.4%) in children. Contrarily, Aslan and Altay (36) concluded that high RBC count cannot be regarded as a reliable preliminary parameter in differentiating IDA from thalassemia in voung children. Okan et al. (37) reported that S&L and G&K indices offered the best discrimination. In Iran, the same contradictory results were reported; Rahim & Keikhaei (38) proved that RBC count and RDWI were the most reliable discrimination indices in patients older than 10 years while S&L index had the highest youden index in those younger than 10 years. Also, Keikhaei (19) concluded that his index, G&K index, RDWI and E&F index are the highest reliable. Similarly, Moghaddam et al. (5) showed that the highest reliability was for Green & King, along with Ricerca, and England & Fraser indices. Batebi et al. (49) revealed that the E&F index had superior sensitivity over S&L index, Mentzer index and traditional red cell indices. Contrarily, Ghafouri et al. (40) recommended Mentzer index. Also. Rajabiani et al. (41) concluded that Mentzer and MDHL achieved the best performance while E&F and S&L was of low sensitivity. In Brazil, high reliability of RDWI was reported by Matos et al. (42) together with G&K index. Also, Lima et al. (43) concluded that G&K index is better than RDW in differentiating IDA from β TT. In contrast to Melo et al. (44) who proposed that βTT was better distinguished from IDA by RBC count and E&F index. Shen et al. (45) studied Chinese children and showed that the highest reliability was for Green & King, along with Ricerca, and England & Fraser indices, while Srivastava and Shine & Lal indices showed the lowest reliability. Ferrara et al. (46) studied Italian children and found that the highest sensitivity was for RDWI, while the highest specificity and Youden index were for E&F index and the highest efficiency was for G&K index. In Egypt, the present study revealed that the highest reliability was for RDWI and Hameed index closely followed by Keikhaei index with the same efficiency. Also, both Hisham and Ricerca indices achieved the next highest accuracy. The differentiating indices of Telmissani, Green & king, Sirdah, differentiating score, England & Fraser and RBCs count moderate reliability gave while Sirachainan et al., Mentzer, Ehsani, Srivastava & Bevington, Huber-Herklotz, MCHD, Bordbar et al, Kerman 1 and Kerman 2 gave low reliability with the least performance was for RDW-SD, Shine & Lal, and Bessman & Feinstein indices (RDW-CV). According to this study, RBCs count alone was not reliable in discriminating between IDA and BTT as IDA in children might be associated with erythrocytosis. It also increased at the initiation of iron therapy and decreased by the end of therapy. A similar observation was made by Rathod et al. (29) who concluded that RBC & RDW and their related formulas (G&K and Ricerca indices) have bad discriminative function compared with cell counter-based parameters particularly MCV and MCH and their related formulas (S&L. Srivastava index, and Mentzer indices) for

the diagnosis of β TT. The role of RDW as parameter an auxiliarv in the differentiation of various anemias is controversial. In some studies (47, 48), a significant difference was observed for RDW between patients with IDA and β TT, while in others this parameter presented low diagnostic accuracy (30, 34, 42, 43, 46, 49-52). However, some studies did not exclude presence of interfering factors or concomitant diseases that could influence RDW performance which is considered a drawback. The four most common mutations found among BTT population; IVS-I-6 (T>C), IVS-I-110 (G>A), IVS-I-1 (G>A) and IVS-II-745 (C>G) followed by other less frequent mutations such as IVS-II-848 (C>A) and codon 5 (-CT), then IVS-II-1 (G>A), -87 (C>G) and codon 39 (C>T) were in concordance with a previous study by Waye et al. (53) who combined data from 5 published surveys of Egyptian β -thalassemia mutations. From the previous data, IVS 1.110 was found to be the most frequent mutation in studies conducted predominantly on βthalassemia major (54-59). Meanwhile, IVS 1.6 was found to be the most frequent studies conducted mutation in predominantly on β thalassemia carriers (with or without β thalassemia intermedia) like this study (60-62). Of all the above literature, this study was the only work to collectively apply twenty four differentiating which indices. when assessed in Egyptian population, the newer discriminant indices e.g. RDWI achieved better performance than the older indices (England and Fraser, Mentzer, Green and King) which may be partly explained by the younger age group as proposed by Hoffmann et al. (63) who demonstrated high variation in the performance of discriminant indices according to age and geographical region, but not the type of hematology analyzer, which considered important factors determining the diagnostic utility of these indices. Also, Hoffmann et al. (63) have objectively shown the superiority of the M/H ratio

over other indices. Despite its high performance, the M/H ratio cannot be used for making a final diagnosis of thalassemia trait. Hussain et al. (64) and Urrechaga et al. (65) proposed formulae that require the availability of a specific generation of hematology analyzers which can give the percentage of hypochromic red cells and the percentage of microcytic red cells and claimed that these formulae were able to efficiently screen patients and confirm The controversies βTT. in relevant literature with different performances for the same index between different regions attributed to different thalassemia is mutations affecting various hematological parameters, degree of severity of IDA, population selection criteria and possibility of concomitant clinical condition. Therefore, larger population based genetic studies correlating between formulae and each of mutations detected in the studied region together with studies proposing new formulae or adopting new cut-offs for existing formulas may prove to be more useful. In conclusion, none of these CBC based indices was completely satisfactory per se in differentiating between BTT or IDA. Studies have shown that these formulas properly identify only 61-91% of the patients with microcytic anemia due to βTT or IDA.

Compliance with ethical standards

Informed consent was taken from participants or from their legal guardians before enrollment into the study.

All procedures performed in this study involving participants were in accordance with the ethical standards approved by the local ethical committee of the Ain Shams University Faculty of Medicine.

Funding

The author did not receive any research grant, this work was funded by the author herself.

Acknowledgement

I would like to express my deep gratitude to Dr. Soha Raouf Youssef, Professor of clinical pathology, Ain Shams University, Cairo, Egypt, for her assistance, great support and comments that greatly improved the manuscript.

Conflict of interest

Author declared no conflict of interst.

References

1. Vehapoglu A, Ozgurhan G, Demir AD, Uzuner S, Nursoy MA, Turkmen S, et al. Hematological indices for differential diagnosis of beta thalassemia trait and iron deficiency anemia. Anemia 2014; 2014:576738-576742.

2. Al Ghwass MM, Halawa EF, Sabry SM, Ahmed D. Iron deficiency anemia in an Egyptian pediatric population: A cross-sectional study. Ann Afr Med 2015;14 (1):25-31.

3. El-Tohamy MK, Eissa AN, El Dorry G, Nasser M, Donia A, Ramzy S. Prevalence of thalassemia carriers among Egyptian school students. Sci Med J ESCME 2003; 15: 4-9.

4. Adlekha S, Chadha T, Jaiswal RM, Singla A. Screening of β -thalassaemia trait by means of red cell indices and derived formulae. Med J Dr D Y Patil Univ 2013; 6 (1):71-74

5. Miri-Moghaddam E, Sargolzaie N. Cut off determination of discrimination indices in differential diagnosis between iron deficiency anemia and β -thalassemia minor. Int J Hematol Oncol Stem Cell Res 2014; 8:1–6.

6. Sachdev R, Dam A, Tyagi G. Detection of Hb variants and hemoglobinopathies in Indian population using HPLC: Report of 2600 cases. Indian J pathol microbial 2010; 53 (1): 57-62.

7. Niazi M, Tahir M, Raziq F, Hameed A. Usefulness of red cell indices in differentiating microcytic hypochromic anemias. Gomal J Med Sci 2010;8:125-129.

8. England JM, Fraser PM. Differentiation of iron deficiency from thalassemia trait by routine blood–count. Lancet 1973;1(7801):447-449.

9. Mentzer WC. Differentiation of iron deficiency from thalassemia trait. Lancet 1973; 1 (7808):882-889.

10. Srivastava PC, Bevington JM. Iron deficiency and–or thalassemia trait. Lancet 1973; 1 (7807):832-835.

11. Shine I, Lal S. A strategy to detect beta thalassemia minor. Lancet 1977; 1(8013):692–694.

12. Green R, King R. A new red blood cell discriminant incorporating volume dispersion for differentiating iron deficiency anemia from thalassemia minor. Blood Cells 1989; 3:481–495.

13. Ricerca BM, Storti S, Onofrio G, Mancini S, Vittori M, Campisi S, et al. Differentiation of iron deficiency from thalassemia trait: A new approach. Haematologica 1987; 72(5):409–413.

14. Bessman JD, Feinstein DI. Quantitative anisocytosis as a discriminant between iron deficiencies and thalassemia minor. Blood 1979;53(2):288–293.

15. Jayabose S, Giamelli J, Levondoglu Tugal O, Sandoval C, Ozkaynak F, Visintainer P. Differentiating iron deficiency anemia from thalassemia minor by using an RDW–based index. J Pediatr Hematol Onclo 1999; 21(4):314-319.

16. Telmissani OA, Khalil S, Roberts GT. Mean density of hemoglobin per liter of blood: a new hematologic parameter with an inherent discriminant function. Lab Hematol 1999; 5:149–152.

17. Ehsani MA; Shahgholi E; Rahiminejad MS; Seighali F; Rashidi A. A new index for discrimination between iron deficiency anemia and beta-thalassemia minor: results in 284 patients. Pak J Biological Sci 2009; 12(5):473-475.

18. Sirdah M, Tarazi I, Al Najjar E, Al Haddad R. Evaluation of the diagnostic reliability of different RBC indices and formulas in the differentiation of the beta– thalassemia minor from iron deficiency in Palestinian population. Int J Lab Hematol 2008; 30 (4): 324–330.

19. Keikhaei B. A new valid formula in differentiating iron deficiency anemia from β-thalassemia trait. Pakist J Med Sci 2010; 26: 368–373. 20. N. P. Sirachainan Iamsirirak Charoenkwan P, Kadegasem P. Wongwerawattanakoon P, Sasanakul W, Chansatitporn N and Chuansumrit A. New Mathematical Formula For Differentiating Thalassemia Trait And Iron Deficiency Anemia In Thalassemia Prevalent Area: A Study In Healthy School-Age Children. Southeast Asian J Trop Med Public Health 2014; 45:1-9.

21. Bordbar E, Taghipour M, Zucconi B. Reliability of Different RBC Indices and Formulas in Discriminating between β -Thalassemia Minor and other Microcytic Hypochromic Cases. Mediterr J Hematol Infect Dis 2015; 7(1): e2015022e2015027.

22. Fairbanks VF, Hines JD, Mazza JJ, Hocking WG. Chapter 2 The Anemias. In: Joseph J Mazza, manual of clinical hematology. Little, brown & co, Boston, New York, Toronto, London 1995;page 17.

23. Huber AR, Ottiger C, Risch L, Regenass S, Hergersberg M, Herklotz R. Thalassämie-syndrome: klinik und diagnose [Syndromes thalassémiques: clinique et diagnostic]. Schweiz Mediz Forum 2004; 4:947–952.

24. Kohan N., Ramzi M. Evaluation of Sensitivity And Specificity Of Kerman Index I And II In Screening Beta Thalassemia Minor. Sci J *Iran* Blood Transfus Org (Khoon) 2008; 4(4) : 297 -302.

25. Hisham A.Getta, Hadeel A. Yasseen, Hameed M. Said. Hi & Ha, are new indices in differentiation between Iron deficiency anemia and beta-Thalassaemia trait /A Study in Sulaimani City-Kurdistan/Iraq. IOSR-JDMS 2015; 14(7):67-72

26. Youden WJ. Index for rating diagnostic tests. Cancer 1950; 3(1):32–35.

27. Šimundić A. Measures of Diagnostic Accuracy: Basic Definitions. EJIFCC 2009; 19(4): 203–211.

28. Nesa A, Abu Tayab , Sultana T, Khondker L, Rahman Q, Anwarulkarim, et al. RDWI is better discriminant than RDW in differentiation of iron deficiency anaemia and β thal trait. Bangladesh J Child Health 2009; 33(3): 100-103.

29. Rathod DA, Kaur A, Patel V, Patel K, Kabrawala R, Patel Vet al. Usefulness of cell counter–based parameters and formulas in detection of beta–thalassemia trait in areas of high prevalence. Am J Clin Pathol 2007;128(4):585–589.

30. Alfadhli SM, Al Awadhi AM, Alkhaldi AD. Validity assessment of nine discriminant functions used for the differentiation between iron deficiency anemia and thalassemia minor. J Trop Pediatr 2007; 53(2):93–97.

31. Urrechaga E. Dicriminant value of % microcytic/%hypochromic ratio in the differential diagnosis of microcytic anemia. Clin Chem Lab Med 2008;46(12):1752–1728.

32. Ntaios G, Chatzinikolaou A, Saouli Z, Girtovitis F, Tsapanidou M, Kaiafa G, et al. Discrimination indices as screening tests for β -thalassemic trait. Ann Hematol 2007;86(7):487–491.

33. Dhara P, Harsh A. Discriminant Functions In Distinguishing Beta Thalassemia Trait and Iron Deficiency Anemia: The value of the RDW-SD. Internet J Hematol 2011; 7 (2): 19-24.

34. Demir A, Yarali N, Fisgin T, Duru F, Kara A: Most reliable indices in differentiation between thalassemia trait and iron deficiency anemia. Pediatr Int 2002; 44(6):612–616.

35. Beyan C, Kaptan K, Ifran A. Predictive value of discrimination indices in differential diagnosis of iron deficiency anemia and beta- thalassemia trait. Eur J Haematol 2007;78(6):524–526.

36. Aslan D, Altay C. Incidence of high erythrocyte count in infants and young children with iron deficiency anemia: re-evaluation of an old parameter. J Pediatric Hematol/Oncol 2003;25(4): 303–306.

37. Okan V, Cigiloglu A, Cifci S, Yilmaz M, Pehlivan M. Red cell indices and functions differentiating patients with the β -thalassaemia trait from those with

DOI: 10.18502/ijpho.v9i3.1163

iron deficiency anaemia. J Int Med Res 2009; 37:25-30.

Rahim F, Keikhaei B, Better 38. differential diagnosis of iron deficiency anemia from beta-thalassemia trait. Turk J Hematol 2009; 26(3):138–145.

39. Batebi A, Pourreza A, Esmailian R. Discrimination of beta-thalassemia minor and iron deficiency anemia by screening test for red blood cell indices. Turk J Med Sci 2012; 42 (2): 275-280.

Ghafouri M, Sefat L, and Sharifi L. 40 Comparison of cell counter indices in differention of beta thalassemia trait and iron deficiency anemia. Scientific J Iranian Blood Transfus Organ 2006; 2(7): 385-389.

41. Rajabiani A, Heshmati P, Ghafouri M. Mean Density of Hemoglobin, a better discriminator of iron deficiency anemia from thalassemia minor. MJIRC 2005; 8: 2-9.

42. Matos JF, Dusse LM, Stubbert RV, Ferreira MR, Coura-Vital W, Fernandes APS et al. Comparison of discriminative indices for iron deficiency anemia and beta thalassemia trait in a Brazilian population. Hematology 2013; 18(3):169-174.

Lima CSP, Reis ARC, Grotto 43. HZW, Saad STO, Costa FF. Comparison of red cell distribution width and a red cell discriminant function incorporating volume dispersion for distinguishing iron deficiency from beta thalassemia trait in patients with microcytosis. São Paulo Med J 1996; 114(5): 1265-1269.

Melo MR, Purini MC, Cancado 44. RD, Kooro F, Chiattone CS. The use of erythrocyte (RBC) indices in the differential diagnosis of microcvtic anemias: is it an approach to be adopted? Rev Ass Med Brasil 2002; 48(3):222-224.

45. Shen C, Jiang YM, Shi H, Liu J, Zhou W, Dai Q, et al. Evaluation of indices in differentiation between iron deficiency anemia and beta-thalassemia trait for Chinese children. J Pediatr Hematol Oncol 2010; 32(6):e218-222.

Ferrara M, Capozzi L, Russo R, 46. Bertocco F, Ferrara D. Reliability of red blood cell indices and formulas to discriminate between beta thalassemia trait children. and iron deficiency in Hematology 2010; 15(2): 112-115.

47. Aslan D, Gumruk F, Gurgey A, Altay C. Importance of RDW value in differential diagnosis of hypochrome anemias. Am J Hematol 2002; 69(1): 31-33.

48. Yermiahu T, Ben-Shalom M, Porath A, Vardi H, Boantza A, Mazor D, et al. Ouantitative determinations of microcytic-hypochromic red blood cell population and glycerol permeability in deficiency anemia iron and beta thalassemia minor. Ann Hematol 1999; 78(10): 468-471.

49. Eldibany MM, Totonchi KF. Joseph NJ, Rhone D. Usefulness of certain red blood cell indices in diagnosing and differentiating thalassemia trait from iron deficiency anemia. Am J Clin Pathol 1999; 111(5): 676-682.

Janel A, Roszyk L, Rapatel C, 50. Mareynat G, Berger MG, Serre-Sapin A. Proposal of a score combining red blood cell indices for early differentiation of beta-thalassemia minor from iron deficiency anemia. Hematology 2011; 16(2): 123-127.

51. Lafferty JD, Crowther MA, Ali MA. The evaluation of various mathematical RBC indices and their efficacy in discriminating between thalassemic and non-thalassemic microcytosis. Am J Clin Pathol 1996; 106(2): 201-215.

Tillyer ML, Tillyer CR. Use of red 52. cell distribution width and erythrocyte zinc protoporphyrin in differential diagnosis of α and β thalassaemia and iron deficiency. J Clin Pathol 1994; 47(3): 205-208.

Waye J, Bory S, Eng B, Patterson 53. M, Chui D, Badr El-Din O, et al. Spectrum of β -thalassemia mutations in Egypt. Hemoglobin 1999; 23 (3): 255-261.

Hussein G, Fawzy M, El-Serafi T, 54. Ismail E, El-Metwally D, Saber M, et al. Rapid detection of β-thalassemia alleles in Egypt using naturally or amplified created

Downloaded from ijpho.ssu.ac.ir on 2025-09-07

DOI: 10.18502/ijpho.v9i3.1163

restriction sites and direct sequencing: A step in disease control. Hemoglobin 2007; 31 (1): 49-62.

55. El-Beshlawy A, Ragab L, Hussein I, El-Tagui M. Genotypes, phenotypes in β -thalassemia and sickle cell anemia in Egypt. JAC 1995; 6(2): 147-151.

56. Omar A, Abdel Karim E, Gendy WE, Marzouk I, Wagdy M. Molecular basis of beta-thalassemia in Alexandria. Egypt J Immunol. 2005; 12 (1): 15-24.

57. Gaafar T, El-Beshlawy A, Aziz M, Abdelrazik H. Rapid screening of β -globin gene mutations by real-time PCR in Egyptian thalassemia children. Afr J Health Sci 2006; 13: 3-4.

58. Naja R, Kaspar H, Shbalko H, Chakar N, Makhoul N, Zalloua P. Accurate and rapid prenatal diagnosis of the most frequent East Mediterranean β thalassemia mutations. Am J Hematol 2004; (75): 220-224.

59. Tadmouri G. Deniz: The electronic database for β -thalassemia mutations in the Arab World. Saudi Med J 2003; 24(24): 1192-1198.

60. Shams El-Din A, Gaafar M, El-Beshlawy A, Sheba H, Ali H. Molecular characterization of non-transfusion dependent β-thalassemia. Egypt J Lab Med 1998; 10 (1): 165-185.

61. Hussein I, El-Beshlawy A, Aboul Ezz E, El-Hadidi S. Screening for β -thalassemia mutations utilizing buccal cells. Egypt J Ped 2000; 17 (2): 139-151.

62. El-Gawhary S, El-Shafie S, Niazi M, Aziz M, El-Beshlawy A. Study of β -thalassemia mutations using the polymerase chain reaction-amplification refractory mutation system and direct sequencing techniques in a group of Egyptian thalassemia patients. Hemoglobin 2007; 31 (1): 63-69.

63. Hoffmann JJ, Urrechaga E, Aguirre U. Meta-analysis of discriminant indices for microcytic anemia. Clin Chem Lab Med 2015; 53(12): 1883–1894.

64. Hussain Z, Malik N, and Chughtai A. Diagnostic Significance of red cell indices in beta thalassemia trait. Biomedica 2005; 21:129-131.

65. Urrechaga E, Borque L, Escanero JF. The role of automated measurement of red cell subpopulations on the Sysmex XE 5000 analyzer in the differential diagnosis of microcytic anemia. Int J Lab Hematol 2011; 33: 30–6.